

ANTIMICROBIAL POTENTIAL OF CINNAMON TAMALA LEAF EXTRACTS

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Abstract: The phytochemical components, and antibacterial activity of extracts of *Cinnamomum tamala* leaves were investigated in this work. Alkaloids, tannins, terpenoids and flavonoids were found to be the most abundant phytochemical elements. The antimicrobial potential of the crude extract and its fractions, namely aqueous, methanolic, n-hexane and chloroform, was examined using the agar well diffusion method against six gram-negative, three gram-positive, and one fungal strain. The extracts assessed exhibited varying degrees of inhibitory zones against all tested microorganisms. The screening reveals that *Cinnamomum tamala* has fascinating therapeutic potential, but more biochemical studies are needed to investigate its putative mode of action.

Keywords: *Cinnamomum tamala*, microbial, fungal

1. INTRODUCTION

Tamale cinnamon (*Buch-Ham.*) Nees and Eberm are tiny evergreen trees of the Lauraceae family. The height of tree is upto 7.5 m with 1.4 circumference (Mishra et al., 2010; Singh et al., 2005) (Kumar et al., 2022). It is found all throughout India because to its tropical and subtropical distribution, although it is most prevalent in the Himalayan area and in Northeast of India. The tree has somewhat rough, dark brown or blackish bark. At 1.3 cm in length, Blaze has whitish streaks that extend towards the outside and a pinkish or reddish brown colour. The leaves might be opposite, sub-opposite, or alternating. They have three nerves that go from just above the base almost to the apex, and they are faintly shiny above ovate, coriaceous, or oblong, lanceolate acuminate. The leaves have several traditional names, including Tamalpatra, Tejpat, Tejpat, and others. They are usually referred to as Indian bay leaves. "Tamalpatra" means "dark leaf" in Sanskrit (Raksha et al., 2021).

The tree's dried leaves are used for making spice. For trees that are 15 years of age or older, a productivity of more than 200 kg on a fresh weight basis is regarded as a satisfactory yield. For five to ten year old trees, it weighs around 100 kg. Starting

of plantation was done with seedlings in Nepal, seeds are gathered from forest, after that self germination is allowed. Typically, harvesting starts around five years old for trees (Aäéiéã et al., 2008).

Due to the presence of numerous major bioactive constituents, the plant has a variety of pharmacological and medicinal properties that can be used to cure and treat a wide range of diseases. These constituents include: myrcene, eugenol (4-hydroxy-3-methoxyallylbenzene), quercetin-3-O-rutinoside, O-glycopyranoside (Astragaloside), camphene, kaempferol-3-O-galloyl, α -pinene, methyl eugenol, p-cymene, limonene, methyl ether eugenol and eugenol acetate. 8–11. The assessment of plant leaf oil revealed that it contains a significant amount of sesquiterpenoids, curcumenol, germacerene D, furanodienone, furanodiene, curzerenone, γ -terpinene, β -caryophyllene, furanogermenone (Haider et al., 2018; Mir et al., 2004).

2. MATERIAL AND METHODS

2.1. CHEMICALS

Methanol, HCl, ethanol, acetic acid, gallic acid, acetic acid (glacial), n-hexane, sulphuric acid, sodium nitrite, sodium hydroxide, aluminium chloride, ferric chloride, quercetin

2.2. COLLECTION AND IDENTIFICATION OF PLANTS

West Bengal, the region Berhampur, was selected for the collection of Fresh *Cinnamomum tamala* leaves. Identification and authentication of *C. tamala* leaves was done from the NISCAIR, New Delhi with letter reference no NISCAIR/RHMD/Consult/2016/2983-10, dated 23/09/2016 (Sharma & Rao, 2014).

3. PHYTOCHEMICAL SCREENING

Various chemical analysis were carried out to determine the presence of secondary active metabolites such as alkaloids, flavonoids, terpenoids, tannins, steroids and saponins (Satyal et al., 2013).

4. CINNAMOMUM TAMALA'S ANTIBACTERIAL TEST AGAINST PARTICULAR BACTERIAL SPECIES

Susceptibility studies were conducted utilizing the modified Agar Well Diffusion method [18] to investigate that the leaves of *Cinnamomum tamala* have antimicrobial properties. The MHA served as the medium. The fixed temperature for incubation was 37°C, and the culture was stored in triplicate for a period of 24 to 72 hours. In a Petri plate, the test organism's broth culture (0.6 mL) was taken. To this the sterilised melted MHA (20mL) was added. Wells were bored and then added (2 mL) of using an extract made from *Cinnamomum tamala* leaves to the medium. An hour was spent on inoculation to guarantee the strong diffusion of antimicrobial agent to the medium (Goyal et al., 2009).

5. ANTI-FUNGAL ASSAY

Agar well diffusion assay was used for antifungal activity. The plant extracts were dissolved in different test tubes containing DMSO. Dextrose agar medium (5mL) was placed in test tube and inoculation was done. Slanting position was selected for keeping of test tube overnight at room temperature. Fungal culture was inoculated on slant. Incubation period was for 7 days at temperature 29°C. Zone of inhibition was calculated (Hassan & Zainab Kazmi, 2015; Satyal et al., 2013).

6. RESULTS

6.1. PHYTOCHEMICAL ANALYSIS

Result of phytochemical screening of various extracts of cinnamon tamala leaves is as follows:

Table I: Phytochemical screening results

Plants	Saponins	Terpenoids	Flavonoides	Steroids	Tannins	Alkaloids
Cinnamomum tamala	-ve	+ve	+ve	-ve	+ve	+ve

6.2. ANTIMICROBIAL ACTIVITY

Antimicrobial potential of *c. tamala* leaves was assayed in vitro by agar well diffusion method. Result is evaluated in table no.2. Variable degree of inhibition was showed by various extracts of *c. tamala* against gram +ve bacterial stains, gram-ve bacterial stains and fungus. The plant extracts showed a significant effectiveness against most of the bacterial and fungal species. Best inhibition activity was shown by all the extracts towards *B. atropheus* but maximum zone of inhibition was shown by aqueous extracts measuring 40 mm. aqueous, methanolic, extracts are completely inactive towards *S. Typhi*.

Table II: Results of antimicrobial activity

Orga nism	Zone of inhibition			Standard drug inhibition		
	n-hex ane	Aqu eous	metha nolic	chloro form	antibi otics	inhibit ion
	12	11	13	14	15	
E coli	11	11	13	13	15	Ciprofl oxacin 36
S. typhi	12	Nil	Nil	Nil	10	Ciprofl oxacin 23
E. cartovo ra	12	13	17	12	11	Ciprofl oxacin 17
A. Tumifa ciens	20	18	19	17	23	Azithro mycin 25

Each author may include his or her biography at the end of the paper. The paragraph begins with the author's name (font size 9 point, bold).

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